Dilutions of the fusion protein test material (50 µl) is added in triplicate to microtiter wells. Test samples (of the protein of interest) are added for final dilution of 1:4; rhuIL-2 (R&D Systems, Minneapolis, MN, catalog number 202-IL) is added to a final concentration of 1 µg/ml; anti-CD4 mAb (R&D Systems, clone 34930.11, catalog number MAB379) is added to a final concentration of 10 µg/ml. Cells are cultured for 7-8 days at 37°C in 5% CO₂, and 1 µC of [³H] thymidine is added to wells for the last 16 hrs of culture. Cells are harvested and thymidine incorporation determined using a Packard TopCount. Data is expressed as the mean and standard deviation of triplicate determinations.

Samples of the fusion protein of interest are screened in separate experiments and compared to the negative control treatment, anti-CD4 mAb, which inhibits proliferation of lymphocytes and the positive control treatment, IL-2 (either as recombinant material or supernatant), which enhances proliferation of lymphocytes.

Example 43: Assays for Protease Activity

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The following assay may be used to assess protease activity of an albumin fusion protein of the invention.

Gelatin and casein zymography are performed essentially as described (Heusen et al., Anal. Biochem., 102:196-202 (1980); Wilson et al., Journal of Urology, 149:653-658 (1993)). Samples are run on 10% polyacryamide/0.1% SDS gels containing 1% gelain orcasein, soaked in 2.5% triton at room temperature for 1 hour, and in 0.1M glycine, pH 8.3 at 37°C 5 to 16 hours. After staining in amido black areas of proteolysis apear as clear areas agains the blue-black background. Trypsin (Sigma T8642) is used as a positive control.

Protease activity is also determined by monitoring the cleavage of n-a-benzoyl-L-arginine ethyl ester (BAEE) (Sigma B-4500. Reactions are set up in (25mMNaPO₄,1mM EDTA, and 1mM BAEE), pH 7.5. Samples are added and the change in adsorbance at 260nm is monitored on the Beckman DU-6 spectrophotometer in the time-drive mode. Trypsin is used as a positive control.

Additional assays based upon the release of acid-soluble peptides from casein or hemoglobin measured as adsorbance at 280 nm or colorimetrically using the Folin method are performed as described in Bergmeyer, et al., *Methods of Enzymatic Analysis*, 5 (1984). Other assays involve the solubilization of chromogenic substrates (Ward, *Applied Science*, 251-317 (1983)).

Example 44: Identifying Serine Protease Substrate Specificity

Methods known in the art or described herein may be used to determine the substrate

specificity of the albumin fusion proteins of the present invention having serine protease activity. A preferred method of determining substrate specificity is by the use of positional scanning synthetic combinatorial libraries as described in GB 2 324 529 (incorporated herein in its entirety).

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Example 45: Ligand Binding Assays

The following assay may be used to assess ligand binding activity of an albumin fusion protein of the invention.

Ligand binding assays provide a direct method for ascertaining receptor pharmacology and are adaptable to a high throughput format. The purified ligand for an albumin fusion protein of the invention is radiolabeled to high specific activity (50-2000 Ci/mmol) for binding studies. A determination is then made that the process of radiolabeling does not diminish the activity of the ligand towards the fusion protein. Assay conditions for buffers, ions, pH and other modulators such as nucleotides are optimized to establish a workable signal to noise ratio for both membrane and whole cell polypeptide sources. For these assays, specific polypeptide binding is defined as total associated radioactivity minus the radioactivity measured in the presence of an excess of unlabeled competing ligand. Where possible, more than one competing ligand is used to define residual nonspecific binding.

Example 46: Functional Assay in Xenopus Oocytes

Capped RNA transcripts from linearized plasmid templates encoding an albumin fusion protein of the invention is synthesized in vitro with RNA polymerases in accordance with standard procedures. In vitro transcripts are suspended in water at a final concentration of 0.2 mg/mi. Ovarian lobes are removed from adult female toads, Stage V defolliculated oocytes are obtained, and RNA transcripts (10 ng/oocyte) are injected in a 50 nl bolus using a microinjection apparatus. Two electrode voltage clamps are used to measure the currents from individual *Xenopus oocytes* in response fusion protein and polypeptide agonist exposure. Recordings are made in Ca2+ free Barth's medium at room temperature. The Xenopus system can be used to screen known ligands and tissue/cell extracts for activating ligands.

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Example 47: Microphysiometric Assays

Activation of a wide variety of secondary messenger systems results in extrusion of small amounts of acid from a cell. The acid formed is largely as a result of the increased metabolic activity required to fuel the intracellular signaling process. The pH changes in the media surrounding the cell are very small but are detectable by the CYTOSENSOR microphysiometer (Molecular Devices Ltd., Menlo Park, Calif.). The CYTOSENSOR is thus

capable of detecting the ability of an albumin fusion protein of the invention to activate secondary messengers that are coupled to an energy utilizing intracellular signaling pathway.

Example 48: Extract/Cell Supernatant Screening

A large number of mammalian receptors exist for which there remains, as yet, no cognate activating ligand (agonist). Thus, active ligands for these receptors may not be included within the ligands banks as identified to date. Accordingly, the albumin fusion proteins of the invention can also be functionally screened (using calcium, cAMP, microphysiometer, oocyte electrophysiology, etc., functional screens) against tissue extracts to identify natural ligands for the Therapeutic protein portion and/or albumin protein portion of an albumin fusion protein of the invention. Extracts that produce positive functional responses can be sequentially subfractionated until an activating ligand is isolated and identified.

Example 49: ATP-binding assay

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The following assay may be used to assess ATP-binding activity of fusion proteins of the invention.

ATP-binding activity of an albumin fusion protein of the invention may be detected using the ATP-binding assay described in U.S. Patent 5,858,719, which is herein incorporated by reference in its entirety. Briefly, ATP-binding to an albumin fusion protein of the invention is measured via photoaffinity labeling with 8-azido-ATP in a competition assay. Reaction mixtures containing 1 mg/ml of ABC transport protein are incubated with varying concentrations of ATP, or the non-hydrolyzable ATP analog adenyl-5'-imidodiphosphate for 10 minutes at 4°C. A mixture of 8-azido-ATP (Sigma Chem. Corp., St. Louis, MO.) plus 8azido-ATP (32P-ATP) (5 mCi/µmol, ICN, Irvine CA.) is added to a final concentration of 100 μM and 0.5 ml aliquots are placed in the wells of a porcelain spot plate on ice. The plate is irradiated using a short wave 254 nm UV lamp at a distance of 2.5 cm from the plate for two one-minute intervals with a one-minute cooling interval in between. The reaction is stopped by addition of dithiothreitol to a final concentration of 2mM. The incubations are subjected to SDS-PAGE electrophoresis, dried, and autoradiographed. Protein bands corresponding to the albumin fusion proteins of the invention are excised, and the radioactivity quantified. A decrease in radioactivity with increasing ATP or adenly-5'-imidodiphosphate provides a measure of ATP affinity to the fusion protein.

Example 50: Phosphorylation Assay

In order to assay for phosphorylation activity of an albumin fusion protein of the invention, a phosphorylation assay as described in U.S. Patent 5,958,405 (which is herein

incorporated by reference) is utilized. Briefly, phosphorylation activity may be measured by phosphorylation of a protein substrate using gamma-labeled ³²P-ATP and quantitation of the incorporated radioactivity using a gamma radioisotope counter. The fusion portein of the invention is incubated with the protein substrate, ³²P-ATP, and a kinase buffer. The ³²P incorporated into the substrate is then separated from free ³²P-ATP by electrophoresis, and the incorporated ³²P is counted and compared to a negative control. Radioactivity counts above the negative control are indicative of phosphorylation activity of the fusion protein.

Example 51: Detection of Phosphorylation Activity (Activation) of an Albumin Fusion Protein of the Invention in the Presence of Polypeptide Ligands

Methods known in the art or described herein may be used to determine the phosphorylation activity of an albumin fusion protein of the invention. A preferred method of determining phosphorylation activity is by the use of the tyrosine phosphorylation assay as described in US 5,817,471 (incorporated herein by reference).

Example 52: Identification Of Signal Transduction Proteins That Interact With An albumin fusion protein Of The Present Invention

Albumin fusion proteins of the invention may serve as research tools for the identification, characterization and purification of signal transduction pathway proteins or receptor proteins. Briefly, a labeled fusion protein of the invention is useful as a reagent for the purification of molecules with which it interacts. In one embodiment of affinity purification, an albumin fusion protein of the invention is covalently coupled to a chromatography column. Cell-free extract derived from putative target cells, such as carcinoma tissues, is passed over the column, and molecules with appropriate affinity bind to the albumin fusion protein. The protein complex is recovered from the column, dissociated, and the recovered molecule subjected to N-terminal protein sequencing. This amino acid sequence is then used to identify the captured molecule or to design degenerate oligonucleotide probes for cloning the relevant gene from an appropriate cDNA library.

Example 53: IL-6 Bioassay

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A variety of assays are known in the art for testing the proliferative effects of an albumin fusion protein of the invention. For example, one such assay is the IL-6 Bioassay as described by Marz et al. (Proc. Natl. Acad. Sci., U.S.A., 95:3251-56 (1998), which is herein incorporated by reference). After 68 hrs. at 37°C, the number of viable cells is measured by adding the tetrazolium salt thiazolyl blue (MTT) and incubating for a further 4 hrs. at 37°C. B9 cells are lysed by SDS and optical density is measured at 570 nm. Controls

containing IL-6 (positive) and no cytokine (negative) are Briefly, IL-6 dependent B9 murine cells are washed three times in IL-6 free medium and plated at a concentration of 5,000 cells per well in 50 μ l, and 50 μ l of fusion protein of the invention is added. utilized. Enhanced proliferation in the test sample(s) (containing an albumin fusion protein of the invention) relative to the negative control is indicative of proliferative effects mediated by the fusion protein.

Example 54: Support of Chicken Embryo Neuron Survival

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To test whether sympathetic neuronal cell viability is supported by an albumin fusion protein of the invention, the chicken embryo neuronal survival assay of Senaldi *et al* may be utilized (*Proc. Natl. Acad. Sci., U.S.A., 96*:11458-63 (1998), which is herein incorporated by reference). Briefly, motor and sympathetic neurons are isolated from chicken embryos, resuspended in L15 medium (with 10% FCS, glucose, sodium selenite, progesterone, conalbumin, putrescine, and insulin; Life Technologies, Rockville, MD.) and Dulbecco's modified Eagles medium [with 10% FCS, glutamine, penicillin, and 25 mM Hepes buffer (pH 7.2); Life Technologies, Rockville, MD.], respectively, and incubated at 37°C in 5% CO₂ in the presence of different concentrations of the purified fusion protein of the invention, as well as a negative control lacking any cytokine. After 3 days, neuron survival is determined by evaluation of cellular morphology, and through the use of the colorimetric assay of Mosmann (Mosmann, T., *J. Immunol. Methods*, 65:55-63 (1983)). Enhanced neuronal cell viability as compared to the controls lacking cytokine is indicative of the ability of the albumin fusion protein to enhance the survival of neuronal cells.

Example 55: Assay for Phosphatase Activity

The following assay may be used to assess serine/threonine phosphatase (PTPase) activity of an albumin fusion protein of the invention.

In order to assay for serine/threonine phosphatase (PTPase) activity, assays can be utilized which are widely known to those skilled in the art. For example, the serine/threonine phosphatase (PSPase) activity of an albumin fusion protein of the invention may be measured using a PSPase assay kit from New England Biolabs, Inc. Myelin basic protein (MyBP), a substrate for PSPase, is phosphorylated on serine and threonine residues with cAMP-dependent Protein Kinase in the presence of [32P]ATP. Protein serine/threonine phosphatase activity is then determined by measuring the release of inorganic phosphate from 32P-labeled MyBP.

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Example 56: Interaction of Serine/Threonine Phosphatases with other Proteins

Fusion protein of the invention having serine/threonine phosphatase activity (e.g., as determined in Example 55) are useful, for example, as research tools for the identification, characterization and purification of additional interacting proteins or receptor proteins, or other signal transduction pathway proteins. Briefly, a labeled fusion protein of the invention is useful as a reagent for the purification of molecules with which it interacts. In one embodiment of affinity purification, an albumin fusion protein of the invention is covalently coupled to a chromatography column. Cell-free extract derived from putative target cells, such as neural or liver cells, is passed over the column, and molecules with appropriate affinity bind to the fusion protein. The fusion protein -complex is recovered from the column, dissociated, and the recovered molecule subjected to N-terminal protein sequencing. This amino acid sequence is then used to identify the captured molecule or to design degenerate oligonucleotide probes for cloning the relevant gene from an appropriate cDNA library.

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Example 57: Assaying for Heparanase Activity

There a numerous assays known in the art that may be employed to assay for heparanase activity of an albumin fusion protein of the invention. In one example, heparanase activity of an albumin fusion protein of the invention, is assayed as described by Vlodavsky et al., (Vlodavsky et al., Nat. Med., 5:793-802 (1999)). Briefly, cell lysates, conditioned media, intact cells (1 x 10^6 cells per 35-mm dish), cell culture supernatant, or purified fusion protein are incubated for 18 hrs at 37°C, pH 6.2-6.6, with 35 S-labeled ECM or soluble ECM derived peak I proteoglycans. The incubation medium is centrifuged and the supernatant is analyzed by gel filtration on a Sepharose CL-6B column (0.9 x 30 cm). Fractions are eluted with PBS and their radioactivity is measured. Degradation fragments of heparan sulfate side chains are eluted from Sepharose 6B at $0.5 < K_{av} < 0.8$ (peak II). Each experiment is done at least three times. Degradation fragments corresponding to "peak II," as described by Vlodavsky et al., is indicative of the activity of an albumin fusion protein of the invention in cleaving heparan sulfate.

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Example 58: Immobilization of biomolecules

This example provides a method for the stabilization of an albumin fusion protein of the invention in non-host cell lipid bilayer constucts (see, e.g., Bieri et al., Nature Biotech 17:1105-1108 (1999), hereby incorporated by reference in its entirety herein) which can be adapted for the study of fusion proteins of the invention in the various functional assays described above. Briefly, carbohydrate-specific chemistry for biotinylation is used to confine a biotin tag to an albumin fusion protein of the invention, thus allowing uniform orientation

upon immobilization. A 50uM solution of an albumin fusion protein of the invention in washed membranes is incubated with 20 mM NaIO4 and 1.5 mg/ml (4mM) BACH or 2 mg/ml (7.5mM) biotin-hydrazide for 1 hr at room temperature (reaction volume, 150ul). Then the sample is dialyzed (Pierce Slidealizer Cassett, 10 kDa cutoff; Pierce Chemical Co., Rockford IL) at 4C first for 5 h, exchanging the buffer after each hour, and finally for 12 h against 500 ml buffer R (0.15 M NaCl, 1 mM MgCl2, 10 mM sodium phosphate, pH7). Just before addition into a cuvette, the sample is diluted 1:5 in buffer ROG50 (Buffer R supplemented with 50 mM octylglucoside).

Example 59: Assays for Metalloproteinase Activity

Metalloproteinases are peptide hydrolases which use metal ions, such as Zn²⁺, as the catalytic mechanism. Metalloproteinase activity of an albumin fusion protein of the present invention can be assayed according to methods known in the art. The following exemplary methods are provided:

Proteolysis of alpha-2-macroglobulin

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To confirm protease activity, a purified fusion protein of the invention is mixed with the substrate alpha-2-macroglobulin (0.2 unit/ml; Boehringer Mannheim, Germany) in 1x assay buffer (50 mM HEPES, pH 7.5, 0.2 M NaCl, 10 mM CaCl₂, 25 μM ZnCl₂ and 0.05% Brij-35) and incubated at 37°C for 1-5 days. Trypsin is used as positive control. Negative controls contain only alpha-2-macroglobulin in assay buffer. The samples are collected and boiled in SDS-PAGE sample buffer containing 5% 2-mercaptoethanol for 5-min, then loaded onto 8% SDS-polyacrylamide gel. After electrophoresis the proteins are visualized by silver staining. Proteolysis is evident by the appearance of lower molecular weight bands as compared to the negative control.

Inhibition of alpha-2-macroglobulin proteolysis by inhibitors of metalloproteinases

Known metalloproteinase inhibitors (metal chelators (EDTA, EGTA, AND $HgCl_2$), peptide metalloproteinase inhibitors (TIMP-1 and TIMP-2), and commercial small molecule MMP inhibitors) may also be used to characterize the proteolytic activity of an albumin fusion protein of the invention. Three synthetic MMP inhibitors that may be used are: MMP inhibitor I, [IC₅₀ = 1.0 μ M against MMP-1 and MMP-8; IC₅₀ = 30 μ M against MMP-9; IC₅₀ = 150 μ M against MMP-3]; MMP-3 (stromelysin-1) inhibitor I [IC₅₀ = 5 μ M against MMP-3], and MMP-3 inhibitor II [K_i = 130 nM against MMP-3]; inhibitors available through Calbiochem, catalog # 444250, 444218, and 444225, respectively). Briefly, different concentrations of the small molecule MMP inhibitors are mixed with a purified fusion protein of the invention (50 μ g/ml) in 22.9 μ l of 1x HEPES buffer (50 mM HEPES, pH 7.5, 0.2 M

NaCl, 10 mM CaCl₂, 25 μM ZnCl₂ and 0.05%Brij-35) and incubated at room temperature (24 °C) for 2-hr, then 7.1 μl of substrate alpha-2-macroglobulin (0.2 unit/ml) is added and incubated at 37°C for 20-hr. The reactions are stopped by adding 4x sample buffer and boiled immediately for 5 minutes. After SDS-PAGE, the protein bands are visualized by silver stain.

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Synthetic Fluorogenic Peptide Substrates Cleavage Assay

The substrate specificity for fusion proteins of the invention with demonstrated metalloproteinase activity may be determined using techniques knonw in the art, such as using synthetic fluorogenic peptide substrates (purchased from BACHEM Bioscience Inc). Test substrates include, M-1985, M-2225, M-2105, M-2110, and M-2255. The first four are MMP substrates and the last one is a substrate of tumor necrosis factor- α (TNF- α) converting enzyme (TACE). These substrastes are preferably prepared in 1:1 dimethyl sulfoxide (DMSO) and water. The stock solutions are 50-500 μ M. Fluorescent assays are performed by using a Perkin Elmer LS 50B luminescence spectrometer equipped with a constant temperature water bath. The excitation λ is 328 nm and the emission λ is 393 nm. Briefly, the assay is carried out by incubating 176 μ l 1x HEPES buffer (0.2 M NaCl, 10 mM CaCl₂, 0.05% Brij-35 and 50 mM HEPES, pH 7.5) with 4 μ l of substrate solution (50 μ M) at 25 °C for 15 minutes, and then adding 20 μ l of a purified fusion protein of the invention into the assay cuvett. The final concentration of substrate is 1 μ M. Initial hydrolysis rates are monitored for 30-min.

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Example 60:Identification and Cloning of VH and VL domains

One method to identfy and clone VH and VL domains from cell lines expressing a particular antibody is to perform PCR with VH and VL specific primers on cDNA made from the antibody expressing cell lines. Briefly, RNA is isolated from the cell lines and used as a template for RT-PCR designed to amplify the VH and VL domains of the antibodies expressed by the EBV cell lines. Cells may be lysed in the TRIzol® reagent (Life Technologies, Rockville. MD) and extracted with one fifth volume of chloroform. After addition of chloroform, the solution is allowed to incubate at room temperature for 10 minutes, and the centrifuged at 14,000 rpm for 15 minutes at 4°C in a tabletop centrifuge. The supernatant is collected and RNA is precipitated using an equal volume of isopropanol. Precipitated RNA is pelleted by centrifuging at 14,000 rpm for 15 minutes at 4°C in a tabletop centrifuge. Following centrifugation, the supernatant is discarded and washed with 75%

ethanol. Follwing washing, the RNA is centrifuged again at 800 rpm for 5 minutes at 4°C. The supernatant is discarded and the pellet allowed to air dry. RNA is the dissolved in DEPC water and heated to 60°C for 10 minutes. Quantities of RNA can determined using optical density measurements.

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cDNA may be synthesized, according to methods well-known in the art, from 1.5-2.5 micrograms of RNA using reverse transciptase and random hexamer primers. cDNA is then used as a template for PCR amplification of VH and VL domains. Primers used to amplify VH and VL genes are shown in Table 3. Typically a PCR reaction makes use of a single 5' primer and a single 3' primer. Sometimes, when the amount of available RNA template is limiting, or for greater efficiency, groups of 5' and/or 3' primers may be used. For example, sometimes all five VH-5' primers and all JH3' primers are used in a single PCR reaction. The PCR reaction is carried out in a 50 microliter volume containing 1X PCR buffer, 2mM of each dNTP, 0.7 units of High Fidelity Taq polymerse, 5' primer mix, 3' primer mix and 7.5 microliters of cDNA. The 5' and 3' primer mix of both VH and VL can be made by pooling together 22 pmole and 28 pmole, respectively, of each of the individual primers. PCR conditions are: 96°C for 5 minutes; followed by 25 cycles of 94°C for 1 minute, 50°C for 1 minute, and 72°C for 1 minute; followed by an extension cycle of 72°C for 10 minutes. After the reaction is completed, sample tubes are stored 4°C.

Table 3: Primer Sequences Used to Amplify VH and VL domains.

	Primer name	SEQ ID NO	Primer Sequence (5'-3')
	VH Primers		
5	Hu VH1-5'	36	CAGGTGCAGCTGGTGCAGTCTGG
10	Hu VH2-5'	37	CAGGTCAACTTAAGGGAGTCTGG
	Hu VH3-5'	38	GAGGTGCAGCTGGTGGAGTCTGG
	Hu VH4-5'	39	CAGGTGCAGCTGCAGGAGTCGGG
	Hu ·VH5-5'	40	GAGGTGCAGCTGTTGCAGTCTGC
	Hu VH6-5'	41	CAGGTACAGCTGCAGCAGTCAGG
	Hu JH1,2-5'	42	TGAGGAGACGGTGACCAGGGTGCC
	Hu JH3-5'	43	TGAAGAGACGGTGACCATTGTCCC
	Hu JH4,5-5'	44	TGAGGAGACGGTGACCAGGGTTCC
	Hu JH6-5'	45	TGAGGAGACGGTGACCGTGGTCCC
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,	VL Primers	*	•
	Hu Vkappa1-5'	46	GACATCCAGATGACCCAGTCTCC
	Hu Vkappa2a-5'	47	GATGTTGTGATGACTCAGTCTCC
	Hu Vkappa2b-5'	48	GATATTGTGATGACTCAGTCTCC
20	Hu Vkappa3-5'	49	GAAATTGTGTTGACGCAGTCTCC
	Hu Vkappa4-5'	50	GACATCGTGATGACCCAGTCTCC
	Hu Vkappa5-5'	51	GAAACGACACTCACGCAGTCTCC
,	Hu Vkappa6-5'	52	GAAATTGTGCTGACTCAGTCTCC
	Hu Vlambda1-5'	53	CAGTCTGTGTTGACGCAGCCGCC
25	Hu Vlambda2-5'	54	CAGTCTGCCCTGACTCAGCCTGC
	Hu Vlambda3-5'	55	TCCTATGTGCTGACTCAGCCACC
	Hu Vlambda3b-5'	56	TCTTCTGAGCTGACTCAGGACCC
	Hu Vlambda4-5'	, 57	CACGTTATACTGACTCAACCGCC
30	Hu Vlambda5-5'	58	CAGGCTGTGCTCACTCAGCCGTC
	Hu Vlambda6-5'	59	AATTTTATGCTGACTCAGCCCCA
	Hu Jkappa1-3'	60	ACGTTTGATTTCCACCTTGGTCCC
	Hu Jkappa2-3'	61	ACGTTTGATCTCCAGCTTGGTCCC
	Hu Jkappa3-3'	62	ACGTTTGATATCCACTTTGGTCCC
	Hu Jkappa4-3'	63	ACGTTTGATCTCCACCTTGGTCCC
35	Hu Jkappa5-3'	64	ACGTTTAATCTCCAGTCGTGTCCC
	Hu Jlambda1-3'	65	CAGTCTGTGTTGACGCAGCCGCC
,	Hu Jlambda2-3'	66	CAGTCTGCCCTGACTCAGCCTGC
40	Hu Jlambda33'	67	TCCTATGTGCTGACTCAGCCACC
	Hu Jlambda3b-3'	68	TCTTCTGAGCTGACTCAGGACCC
	Hu Jlambda4-3'	69	CACGTTATACTGACTCAACCGCC
	Hu Jlambda5-3'	70	CAGGCTGTGCTCACTCAGCCGTC
	Hu Jlambda6-3'	71	AATTTTATGCTGACTCAGCCCCA

PCR samples are then electrophoresed on a 1.3% agarose gel. DNA bands of the expected sizes (~506 base pairs for VH domains, and 344 base pairs for VL domains) can be cut out of the gel and purified using methods well known in the art. Purified PCR products can be ligated into a PCR cloning vector (TA vector from Invitrogen Inc., Carlsbad, CA). Individual cloned PCR products can be isolated after transfection of E. coli and blue/white color selection. Cloned PCR products may then be sequenced using methods commonly known in the art.

The PCR bands containing the VH domain and the VL domains can also be used to create full-length Ig expression vectors. VH and VL domains can be cloned into vectors containing the nucleotide sequences of a heavy (e.g., human IgG1 or human IgG4) or light chain (human kappa or human lambda) constant regions such that a complete heavy or light chain molecule could be expressed from these vectors when transfected into an appropriate host cell. Further, when cloned heavy and light chains are both expressed in one cell line (from either one or two vectors), they can assemble into a complete functional antibody molecule that is secreted into the cell culture medium. Methods using polynucleotides encoding VH and VL antibody domain to generate expression vectors that encode complete antibody molecules are well known within the art.

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It will be clear that the invention may be practiced otherwise than as particularly described in the foregoing description and examples. Numerous modifications and variations of the present invention are possible in light of the above teachings and, therefore, are within the scope of the appended claims.

The entire disclosure of each document cited (including patents, patent applications, patent publications, journal articles, abstracts, laboratory manuals, books, or other disclosures) as well as information available through Identifiers specific to databases such as GenBank, GeneSeq, or the CAS Registry, referred to in this application are herein incorporated by reference in their entirety. The specification and sequence listing of each of the following U.S. applications are herein incorporated by reference in their entirety: Application Nos. 09/091,873 filed June 25, 1998; 60/229,358 filed on April 12, 2000; 60/199,384 filed on April 25,2000; 60/256,931 filed on December 21, 2000, 09/809,269, filed March 16, 2001; 60/277,980, filed March 23, 2001; 09/236,557, filed January 26, 1999; 09/482,273, filed January 13, 2000; 60/234,925, filed November 1, 2000; 09/397,945, filed September 17, 1999; 09/296,622, filed April 23, 1999; 60/092,921, filed July 15, 1998; 09/305,736, filed May 5, 1999; 09/781,417, filed February 13, 2001; 60/152,317, filed September 3, 1999; 09/227,357, filed January 8, 1999; and 60/262,066,

filed January 18, 2001; and International Publication Nos. WO98/39446, filed September 11, 1998; WO 00/61625, filed October 19, 2000; WO/00/77022, filed December 21, 2000; and WO/00/76530, filed December 21, 2000.

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A. The indications made below relate to the deposited microorganism or other biological material referred to in the description on page 96, line 30.					
B. IDENTIFICATION OF DEPOSIT	Further deposits are identified on an additional sheet				
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Address of depositary institution (including postal 10801 University Boulevard Manassas, Virginia 20110-2209 United States of America	code and country)				
Date of deposit	Accession Number				
11 April 2001	Unassigned				
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ATCC Deposit No.: Unassigned

DENMARK

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SWEDEN

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NETHERLANDS

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OR OTHER BIOLOGICAL MATERIAL (PCT Rule 13bis)					
B. IDENTIFICATION OF DEPOSIT	Further deposits are identified on an additional sheet				
Name of depositary institution: American Type C	ulture Collection				
Address of depositary institution (including postal 10801 University Boulevard Manassas, Virginia 20110-2209 United States of America	code and country)				
Date of deposit	Accession Number				
11 April 2001	Unassigned				
C. ADDITIONAL INDICATIONS (leave blank if not applied	cable) This information is continued on an additional sheet				
D. DESIGNATED STATES FOR WHICH INDICATIO	NS ARE MADE (if the indications are not for all designated States)				
Europe In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC). Continued on additional sheets					
E. SEPARATE FURNISHING OF INDICATIONS (leave	blank if not applicable)				
The indications listed below will be submitted to the international I Number of Deposit")	Bureau later (specify the general nature of the indications e.g., "Accession				
For receiving Office use only	For International Bureau use only				
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Revised Form PCT/RO/134 (January 2001)

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CANADA

The applicant requests that, until either a Canadian patent has been issued on the basis of an application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the Commissioner of Patents only authorizes the furnishing of a sample of the deposited biological material referred to in the application to an independent expert nominated by the Commissioner, the applicant must, by a written statement, inform the International Bureau accordingly before completion of technical preparations for publication of the international application.

NORWAY

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AUSTRALIA

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FINLAND

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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM OR OTHER BIOLOGICAL MATERIAL

	OTHERDIC	DOGICAL III	AIBRIAL		
(PCT Rule 13bis)					
A. The indications made below relate to the deposited microorganism or other biological material referred to in the description on page 96, line 30.					
B. IDENTIFICATION OF DEPOSIT		Further deposits are identified on an additional sheet			
Name of depositary institution: American Type Culture Collection					
Address of depositary institution <i>(incli</i> 10801 University Boulevard Manassas, Virginia 20110-2209 United States of America	uding postal	code and co	untry)		
Date of deposit		Accession Number			
11 April 2001		Unassigned			
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet					
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)					
Europe In respect of those designations in which a European Patent is sought a sample of the deposited microorganism will be made available until the publication of the mention of the grant of the European patent or until the date on which the application has been refused or withdrawn or is deemed to be withdrawn, only by the issue of such a sample to an expert nominated by the person requesting the sample (Rule 28(4) EPC). Continued on additional sheets					
E. SEPARATE FURNISHING OF INDICATIONS (leave blank if not applicable)					
The indications listed below will be submitted to the international Bureau later (specify the general nature of the indications e.g., "Accession Number of Deposit")					
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ATCC Deposit No.: Unassigned

CANADA

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What is claimed:

1. An albumin fusion protein comprising a Therapeutic protein:X and albumin comprising the amino acid sequence of SEQ ID NO:18.

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- 2. An albumin fusion protein comprising a Therapeutic protein:X and a fragment or a variant of the amino acid sequence of SEQ ID NO:18, wherein said fragment or variant has albumin activity.
- 3. The albumin fusion protein of claim 2, wherein said albumin activity is the ability to prolong the shelf life of the Therapeutic protein:X compared to the shelf-life of the Therapeutic protein:X in an unfused state.
- 4. The albumin fusion protein of claim 2, wherein the fragment or variant comprises the amino acid sequence of amino acids 1-387 of SEQ ID NO:18.
 - 5. An albumin fusion protein comprising a fragment or variant of a Therapeutic protein:X, and albumin comprising the amino acid sequence of SEQ ID NO:18, wherein said fragment or variant has a biological activity of the Therapeutic protein:X.

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- 6. The albumin fusion protein of any one of claims 1-5, wherein the Therapeutic protein:X, or fragment or variant thereof, is fused to the N-terminus of albumin, or the N-terminus of the fragment or variant of albumin.
- 7. The albumin fusion protein of any one of claims 1-5, wherein the Therapeutic protein:X, or fragment or variant thereof, is fused to the C-terminus of albumin, or the C-terminus of the fragment or variant of albumin.
 - 8. The albumin fusion protein of any one of claims 1-5, wherein the

Therapeutic protein:X, or fragment or variant thereof, is fused to the N- terminus and C-terminus of albumin, or the N-terminus and the C-terminus of the fragment or variant of albumin.

5 9. The albumin fusion protein of any one of claims 1-5, which comprises a first Therapeutic protein:X, or fragment or variant thereof, and a second Therapeutic protein:X, or fragment or variant thereof, wherein said first Therapeutic protein:X, or fragment or variant thereof, is different from said second Therapeutic protein:X, or fragment or variant thereof.

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- 10. The albumin fusion protein of any one of claims 1-8, wherein the Therapeutic protein:X, or fragment or variant thereof, is separated from the albumin or the fragment or variant of albumin by a linker.
- 15 11. The albumin fusion protein of any one of claims 1-8, wherein the albumin fusion protein has the following formula:

R1-L-R2; R2-L-R1; or R1-L-R2-L-R1,

wherein R1 is Therapeutic protein:X, or fragment or variant thereof, L is a peptide linker, and R2 is albumin comprising the amino acid sequence of SEQ ID NO:18 or fragment or variant of albumin.

12. The albumin fusion protein of any one of claims 1-11, wherein the shelf-life of the albumin fusion protein is greater than the shelf-life of the Therapeutic protein:X in an unfused state.

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13. The albumin fusion protein of any one of claims 1-11, wherein the in vitro biological activity of the Therapeutic protein:X, or fragment or variant thereof, fused to albumin, or fragment or variant thereof, is greater than the in vitro biological activity of the Therapeutic protein:X, or a fragment or variant thereof, in an unfused state.

14. The albumin fusion protein of any one of claims 1-11, wherein the in vivo biological activity of the Therapeutic protein:X, or fragment or variant thereof, fused to albumin, or fragment or variant thereof, is greater than the in vivo biological activity of the Therapeutic protein:X, or a fragment or variant thereof, in an unfused state.

15. An albumin fusion protein comprising a Therapeutic protein:X, or fragment or variant thereof, inserted into an albumin comprising the amino acid sequence of SEQ ID NO:18 or fragment or variant thereof.

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- 16. An albumin fusion protein comprising a Therapeutic protein:X, or fragment or variant thereof, inserted into an albumin comprising an amino acid sequence selected from the group consisting of:
 - (a) amino acids 54 to 61 of SEQ ID NO:18;
- (b) amino acids 76 to 89 of SEQ ID NO:18;
 - (c) amino acids 92 to 100 of SEQ ID NO:18;
 - (d) amino acids 170 to 176 of SEQ ID NO:18;
 - (e) amino acids 247 to 252 of SEQ ID NO:18;
 - (f) amino acids 266 to 277 of SEQ ID NO:18;
 - (g) amino acids 280 to 288 of SEQ ID NO:18;
 - (h) amino acids 362 to 368 of SEQ ID NO:18;
 - (i) amino acids 439 to 447 of SEQ ID NO:18;
 - (j) amino acids 462 to 475 of SEQ ID NO:18;
 - (k) amino acids 478 to 486 of SEQ ID NO:18; and
- 25 (1) amino acids 560 to 566 of SEQ ID NO:18.
 - 17. The albumin fusion protein of claims 15 or 16, wherein said albumin fusion protein comprises a portion of albumin sufficient to prolong the shelf-life of the Therapeutic protein:X, or fragment or variant thereof, as compared to the shelf-life of the Therapeutic

protein:X, or a fragment or variant thereof, in an unfused state.

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18. The albumin fusion protein of claims 15 or 16, wherein said albumin fusion protein comprises a portion of albumin sufficient to prolong the in vitro biological activity of the Therapeutic protein:X, or fragment or variant thereof, fused to albumin as compared to the in vitro biological activity of the Therapeutic protein:X, or a fragment or variant thereof, in an unfused state.

- 19. The albumin fusion protein of claims 15 or 16 wherein said albumin fusion protein comprises a portion of albumin sufficient to prolong the in vivo biological activity of the Therapeutic protein:X, or fragment or variant thereof, fused to albumin compared to the in vivo biological activity of the Therapeutic protein:X, or a fragment or variant thereof, in an unfused state.
- 15 20. The albumin fusion protein of any one of claims 1-19, which is non-glycosylated.
 - 21. The albumin fusion protein of any one of claims 1-19, which is expressed in yeast.
 - 22. The albumin fusion protein of claim 21, wherein the yeast is glycosylation deficient.
- 23. The albumin fusion protein of claim 21 wherein the yeast is glycosylation and protease deficient.
 - 24. The albumin fusion protein of any one of claims 1-19, which is expressed by a mammalian cell.

25. The albumin fusion protein of any one of claims 1-19, wherein the albumin fusion protein is expressed by a mammalian cell in culture.

- The albumin fusion protein of any one of claims 1-19, wherein the albumin
 fusion protein further comprises a secretion leader sequence.
 - 27. A composition comprising the albumin fusion protein of any one of claims 1-26 and a pharmaceutically acceptable carrier.
- 10 28. A kit comprising the composition of claim 27.
 - 29. A method of treating a disease or disorder in a patient, comprising the step of administering the albumin fusion protein of any one of claims 1-26.
- The method of claim 29, wherein the disease or disorder comprises indication: Y.
- 31. A method of treating a patient with a disease or disorder that is modulated by Therapeutic protein:X, or fragment or variant thereof, comprising the step of administering an effective amount of the albumin fusion protein of any one of claims 1-26.
 - 32. The method of claim 31, wherein the disease or disorder is indication. Y.
- 33. A method of extending the shelf life of Therapeutic protein:X comprising the step of fusing the Therapeutic protein:X, or fragment or variant thereof, to albumin or a fragment or variant thereof, sufficient to extend the shelf-life of the Therapeutic protein:X, or fragment or variant thereof, compared to the shelf-life of the Therapeutic protein:X, or a fragment or variant thereof, in an unfused state.

34. A nucleic acid molecule comprising a polynucleotide sequence encoding the albumin fusion protein of any one of claims 1-26.

35. A vector comprising the nucleic acid molecule of claim 34.

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36. A host cell comprising the nucleic acid molecule of claim 35.

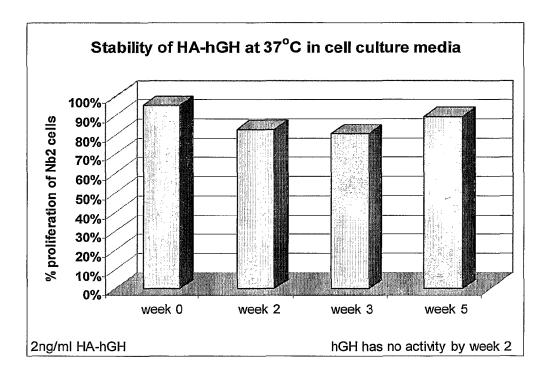


Figure 1

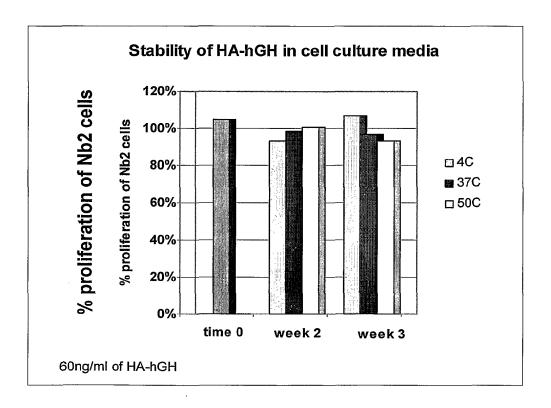


Figure 2

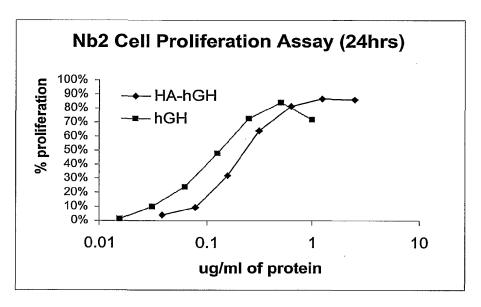


Figure 3A

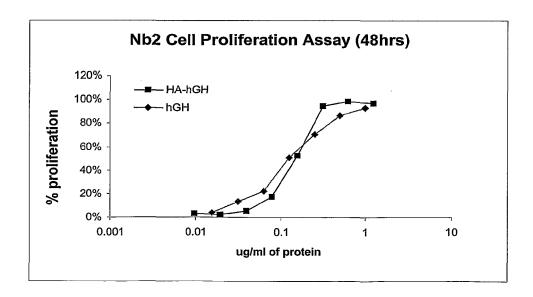


Figure 3B



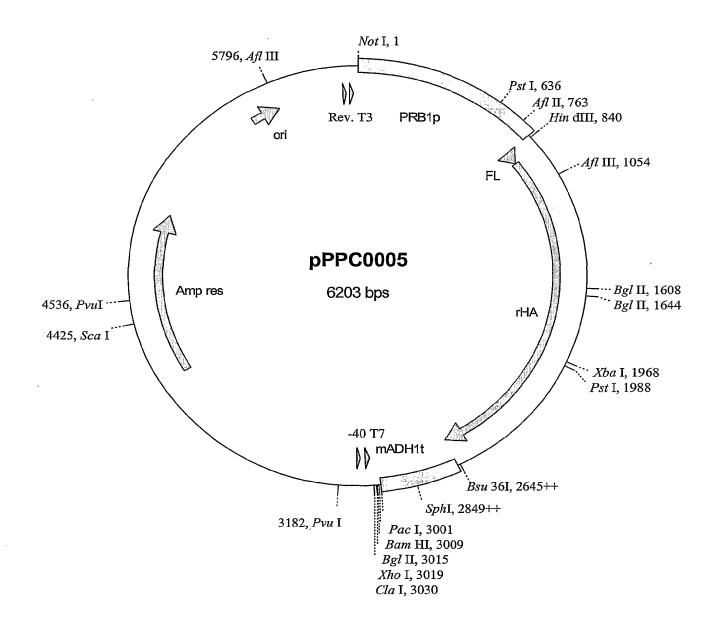


Figure 4

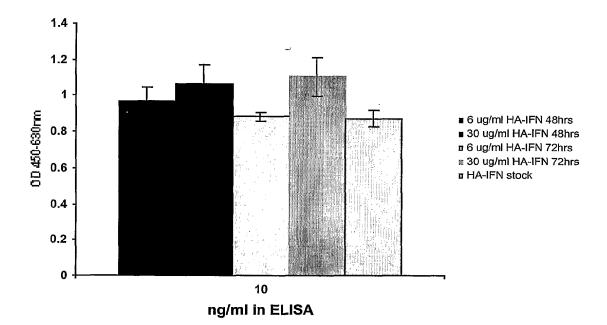
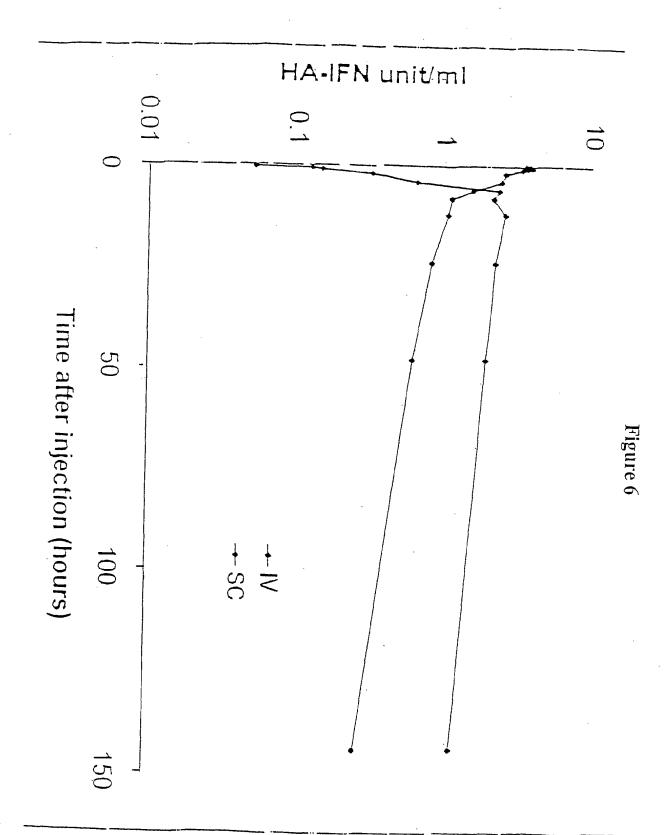


Figure 5



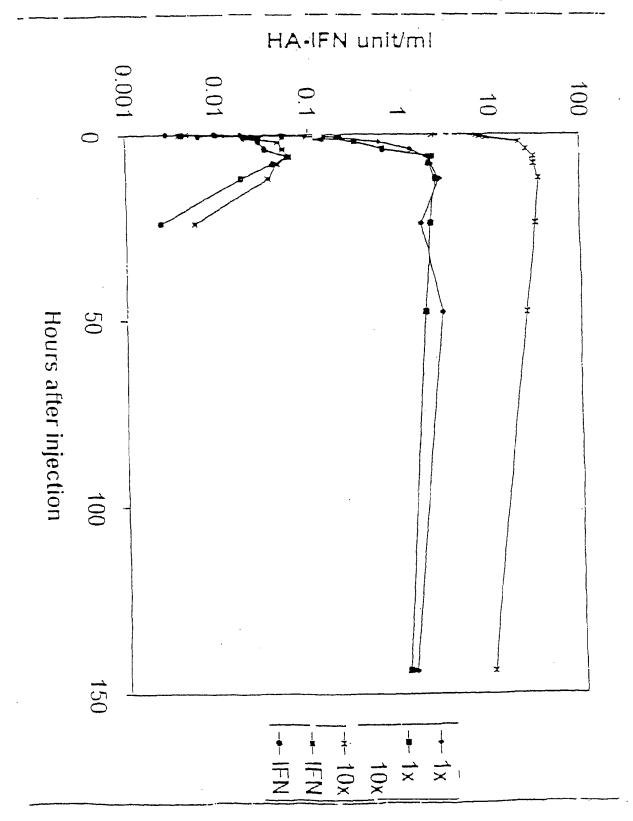
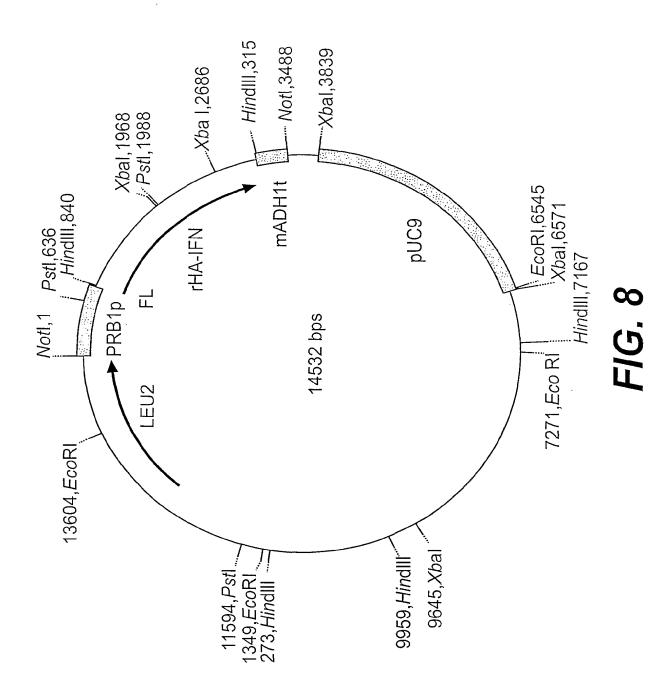


Figure 7

8/20



9/20

<u>Localisation of 'Loops' based on the HA Crystal Structure</u> which could be used for Mutation/Insertion

1	DAHKSEVAHR HHHHH	FKDLGEENFK HHH HHH	ALVLIAFAQY ННННННННН	LQQCPFEDHV HHHHH		
51	I KTCV ADESAE HHHHH		GDKLC TVATL ННННН	II RETYGEMADC HHHH		
101	CFLQHKDDNP HHHH			EETFLKKYLY HHHHHHHHH		
151	APELLFFAKR ННННННННН	ҮКААГТЕСС <u>о</u> нинининин	TV AADKAACLLP ННННН	KLDELRDEGK НННЕННННН	ASSAKQRLKC НИННИННИН	
201				VSKLVTDLTK ННННННННН		
		v	'I	VII	•	
251	LE CADDRADL НННННННННН	AKYIC ENODS	_	KPLLEKSHCI HHHHHHH		
301	DLPSLAADFV HHHH	ESKDVCKNYA HHHHHH		LYEYARRHPD HHHHHH		
351	КТҮЕТТЬЕКС НИНИНИННН			VEEPQNLIKQ ННННННННН		
401		YTKKVPQVST HHHH H		GKVGSKCC KH HHH	IX PEAKRMP CAE НИННИНН	
4==		х		ХI		
451	DYLSVVLNQL ННННННННН			LVNRRPPCFSA HHHHHHHH	A LEVDETYVPK	
501	EFNAETFTFH		RQIKKQTALV HHHHMMEHHH	ELVKHKPKAT HHH	KEQLKAVMDD НННННННН	
551	FAAFVEKCC <u>K</u> НННННННН	XII <u>ADDKET</u> CFAE HHHH	EGKKLVAASQ ННИННИННН			
	II Thr7 III Ala9 IV Gln1 V His2	4-Asn61 6-Asp89 2-Glu100 70-Ala176 47-Glu252 66-Glu277	Loop VII VIII IX X XI XII	Glu280-His: Ala362-Glu: Lys439-Pro- Val462-Lys- Thr478-Pro- Lys560-Thr:	368 447 475 486	

Figure 9

SUBSTITUTE SHEET (RULE 26)

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Examples of Modifications to Loop IV

a. Randomisation of Loop IV.

IV

ΙV

X represents the mutation of the natural amino acid to any other amino acid. One, more or all of the amino acids can be changed in this manner. This figure indicates all the residues have been changed.

b. Insertion (or replacement) of Randomised sequence into Loop IV.



The insertion can be at any point on the loop and the length a length where n would typically be 6, 8, 12, 20 or 25.

Figure 10

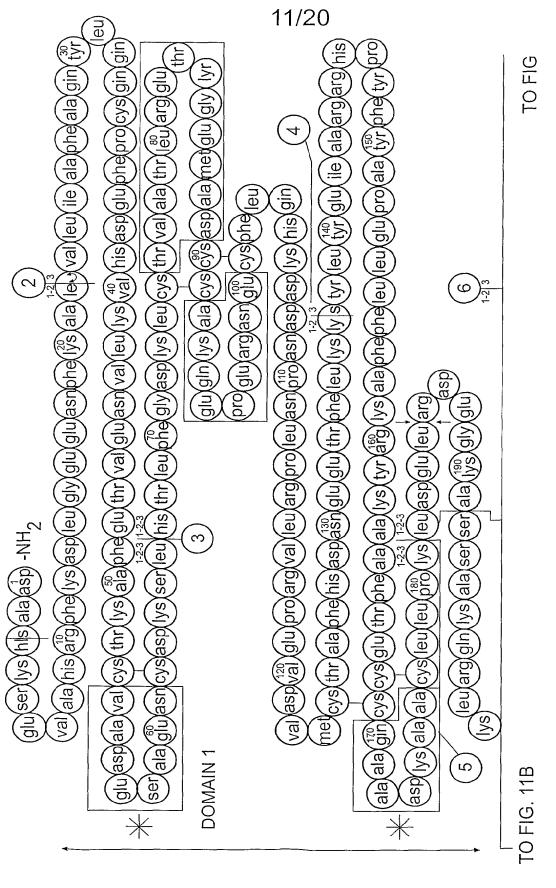
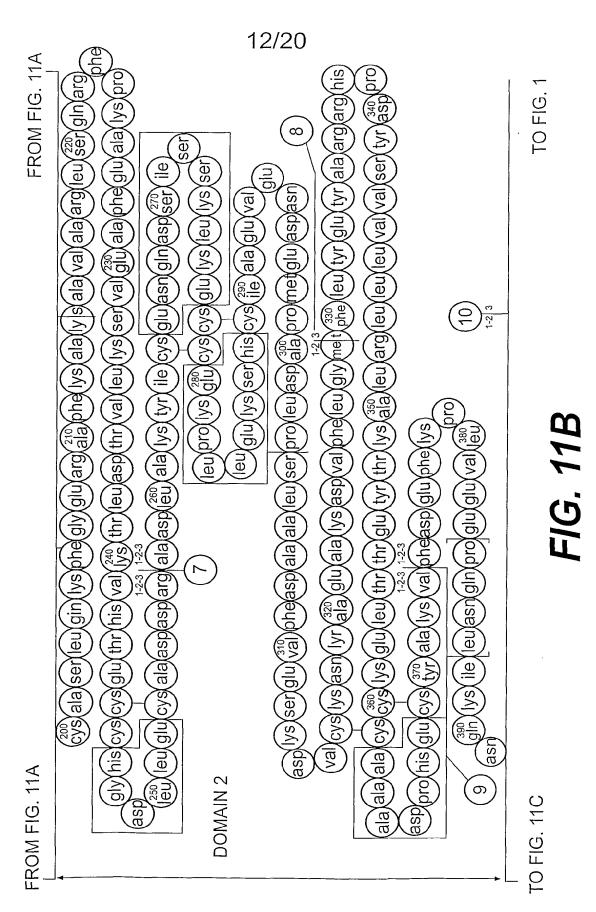


FIG. 11A



SUBSTITUTE SHEET (RULE 26)

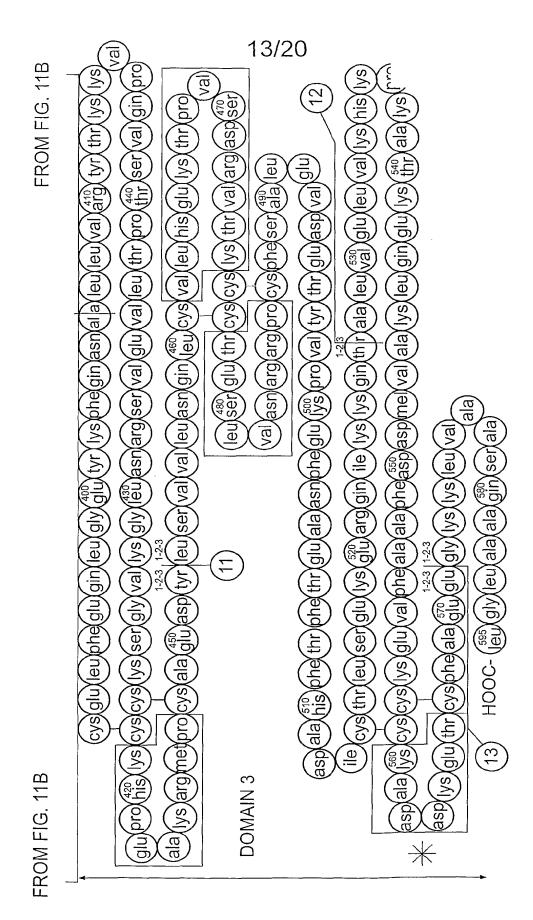
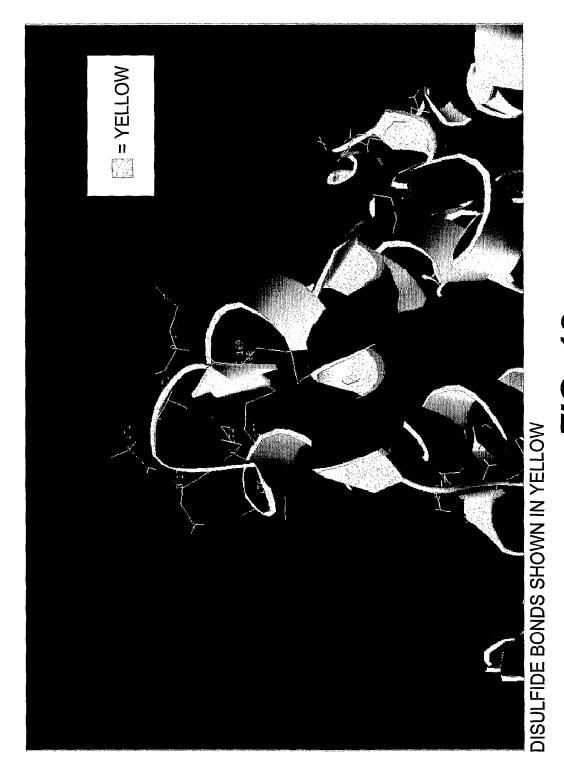


FIG. 11C

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F/G. 12: LOOP IV GLU170-A176

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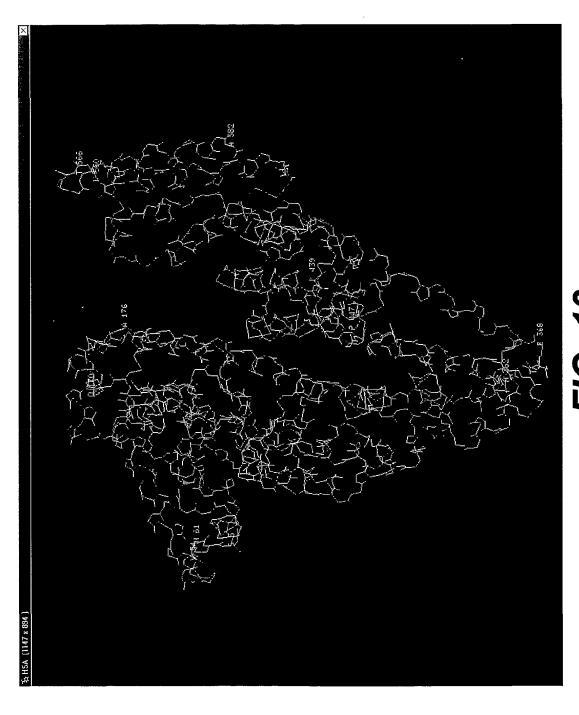
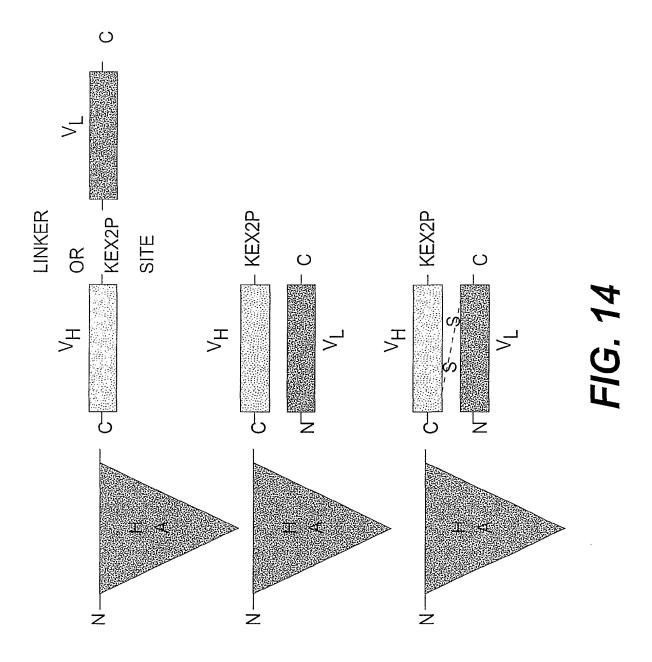


FIG. 13 TERTIARY STRUCTURE OF HA

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60	120	180	240 80	300	360 120	420 140	480
AAA K	GTA V	GAA	CTT L	Gaa e	GTT	TAT Y	AGG R
${ m TTC}$	CAT H	GCT A	ACT T	AAT N	GAG E	TTA L	aaa K
AAT N	GAT D	TCA S	GCA	AGA R	CCA P	TAC Y	GCT
GAA E	GAA E	GAG E	GTT V	GAG E	AGA R	AAA K	TTT F
gaa E	TTT F	GAT D	ACA T	CCT	GTG V	AAA K	TTC F
GGA	CCA P	GCT A	TGC	GAA E	$\mathop{\mathrm{TTG}}_{\mathrm{L}}$	$ ext{TTG}$	$_{\rm L}^{\rm CTT}$
TTG L	TGT	GTT V	TTA L	CAA Q	· CGA R	TTT F	CTC CTT L L
GAT D	CAG Q	TGT	AAA K	AAA K	CCC DDC	ACA T	gaa E
AAA K	CAG Q	ACA T	GAC D	GCA A	CTC L	GAG E	CCG
TTT F	$_{\rm L}^{\rm CTT}$	AAA K	GGA GAC G D	TGT GCA C	AAC CTC (N L	GAA E	GCC CCG A P
CGG R	$_{\rm TAT}^{\cdot}$	GCA A	TTT F	TGC C	CCA P	AAT N	TAT Y
CAT H	CAG Q	ľŢŢ	$_{\rm L}^{\rm CTT}$	GAC D	AAC N	GAC D	TTT F
GCT A	GCT A	GAA '	ACC	GCT GAC TGC 1 A D C	GAC AAC CCA P D N P N	CAT H	TAC TTT '
GTT V	TTT F	ACT T	CAT H	ATG M	GAT D	TTT F	CCT P
GAG E	GCC	GTA V	CTT L	GAA E	aaa K	GCT	CAT H
AGT S	ATT I	GAA E	TCA S	$_{\rm GGT}$	CAC H	ACT T	AGA R
AAG K	$_{\rm TTG}^{\cdot}$	AAT N	AAA K	TAT Y	CAA Q	TGC	AGA R
CAC H	GTG V	GTG V	GAC D	ACC T	${\rm TTG}_{\rm L}$	ATG M	GCC A
GCA A	${ m TTG}$	· TTA L	TGT	GAA E	${ m TTC}$	GTG V	ATT I
GAT D	GCC	AAA K	AAT '	CGT R	TGC	GAT D	GAA E
	61 21	121	181 61	241 81	301	361	421 141

figure 15A

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540 180	600	660	720	780 260	840 280	300	960 320
CCA P	TGT C	AGC S	ACC AAA 7 T K 2	CTT L	GAA E	GAG ATG CCT GCT E M P A	GAC TTG CCT TCA TTA GCT GCT GAT TTT GTT GAA AGT AAG GAT GTT TGC AAA AAC TAT GCT D L P S L A A D F V E S K D V C K N Y A
$ ext{TTG}$	AAA K	CTG	ACC	GCT GAT GAC AGG GCG GAC A D D R A D	GAA TGC TGT (CCT	TAT Y
CTG L	CTC	CGC R	CTT	GCG A	TGC	ATG M	AAC N
TGC CTG	AGA CTC I R L	GCT CGC (GAT CTT 1 D L	AGG R	GAA E	GAG E	AAA K
AAA GCT GCC 1 K A A C	GCC AAA CAG 1 A K Q 1	TGG GCA GTG (TTA GTG ACA (GAC D	AAA CTG AAG C K L K I	GAT D	TGC
GCT A	AAA K	GCA A	GTG V	GAT D	${ m crg}_{ m L}$	AAT N	GTT
AAA K	GCC A	TGG W	$_{\rm TTA}^{\cdot}$	GCT A	AAA K	GAA E	GAT D
GAT D	TCT S	TTC AAA GCA 3 F K A 1	TCC AAG	${ m TGT}$	AGT	GTG V	AAG K
GCT A	TCG	AAA K	TCC	GAA E	TCC	gaa E	AGT S
GCT A	GCT A	TTC F	$_{\rm V}^{\rm GTT}$	CTT L	ATC I	GCC A	GAA E
CAA Q	AAG K	GCT A	TTT GCA GAA GTT F A E V	CTG L	TCG S	ATT I	GTT V
TGC	999	AGA R	GCA A	GAT D	GAT D	TGC	TTT F
TGT C	gaa E	GAA E	TTT F	GGA G	CAG Q	CAC H	GAT D
GAA E	GAT	GGA G	GAG E	CAT H	AAT N	TCC S	GCT A
TTT ACA GAA TGT TGC CAA GCT GCT GAT	CTT CGG GAT GAA GGG AAG GCT TCG L R D E G K A S	CTC CAA AAA TTT GGA GAA AGA GCT L Q K F G E R A	AAA GCT GAG 1 K A E F	TGC TGC CAT GGA GAT CTG CTT GAA TGT C C H G D L L E C	ATC TGT GAA AAT CAG GAT TCG ATC TCC AGT	GAA AAA TCC CAC TGC ATT GCC GAA GTG GAA AAT GAT E K S H C I A E V E N D	GCT A
TTT F	CTT L	AAA K	aaa K	TGC	TGT	GAA E	TTA L
AAA GCT GCT 1 K A A F	GAA E·	CAA Q	CCC P	GAA E	ATC I	CTG TTG (L L	TCA S
GCT A	GAT D	CTC	TTT F	ACG	TAT	$_{ m L}$	CCT
AAA K	CTC L	AGT S	AGA R	CAC H	AAG K	CCT P	${ m TTG}$
TAT Y	AAG K	GCC	CAG Q	GTC V	GCC AAG 7 A K	AAA CCT (K P]	GAC D
481 161	541 181	601	661 221	721 241	781 261	841 281	901 301

Figure 15B

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GAT GTC TTC CTG GGC ATG TTT TTG TAT GAA TAT GCA AGA AGG CAT CCT GAT 1020 D V F L G M F L Y E Y A R R H P D 340	A GAG AAG TGC 1080 E K C 360	F AAA CCT CTT 1140 K P L 380		S CTT GGA GAG 1200 L G E 400	CTT GGA GAG L G E . GTG TCA ACT V S T
A AGG R	I CTA	TTT F		CCT CAG AAT TTA ATC AAA CAA AAC TGT GAG CTT TTT GAG CAG P Q N L I K Q N C E L F E Q	G CAA
3CA AG	CTG AGA CTT GCC AAG ACA TAT GAA ACC ACT L R L A K T Y E T T	TAT GCC AAA GTG TTC GAT GAA TTT Y A K V F D E F		rtt GA(TTT GAG F E GTA CCC
TAT (GAA 1	TTC (CTT	L F F · · · · · · · · · · · · · · · · ·
IT GAA E	'A TAT Y	AA GTG V	T GAG	臼	E C AAG K
rrg ra	aag ac K T	3CC AA	AAC TG	D D	Q N L I K Q N C E
TTT 1 F I	GCC 7	TAT (Y	CAA A		CGT T
ATG M	A CTT L	CAT GAA TGC 1 H E C Y	AAA K		A GTT
9 9	G AGA R	T GAA E	A ATC I		A TTA L
ic cr	G CTC L	CT CA	T. T.		IG CT7
TC TT	TG CT	AT CC	AG AA N		AT GC
GAT G D V	GIG CTG CTG C	GCA GAT CCT C A D P F	CCT C P Q		CAG A Q N
GAG GCA AAG G E A K L	GTC V	GCT A	GAA GAG (E E]		TTC F
GCA A	TCT S	GCC	GAA E		AAA K
LGAG ∪E	1 TAC	1 TGT 1 C	L GTG L V		L TĄC L Y
961 321	1021 341	1081 361	1141 381		1201

Figure 15(

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1500 500	1560 520	1620 540	1680 560	1740 580	
aaa K	GAG E	ACA T	AAG K	CAA Q	
CCC P	AAG. K	GCA A	TGC	AGT S	
GTT V	GAG E	AAG K	TGC	GCA A	
${ m TAC}$	TCT	CCC P	AAG K	GTT GCT V A	
ACA T	$_{\rm L}^{\rm CTT}$	AAG K	GAG E		0.1
GAA E	ACA T	CAC H	GTA V	AAA AAA CTT K K L	1782 585
GAT D	TGC C	AAA K	TTT F	AAA K	CAG
GTC V	ATA I	GTG AAA CAC AAG CCC V K H K P	GCT TTT GTA A F V	AAA K	TCT
	GAT D	$_{\rm L}^{\rm CTT}$	GCA A	GGT	GCA
CTG L	CAT GCA H A	GAG E	${ m TTC}$	GAG E	TTA AAA
GCT CTG A L	CAT H	CTT GTT GAG L V E	GAT TTC D	GAG GAG (E E (TTA
TCA S	TTC	CTT L	GAT D	GCC A	CAT
TGC TTT C F	ACC T	ATC AAG AAA CAA ACT GCA I K K Q T A	ATG M	ACC TGC TTT T C F	CTA
TGC	TTC F	ACT T	GTT V	TGC C	CAT
CCA P	ACA T	CAA Q	GCT A	ACC	TTA GGC TTA TAA L G L *
CGA R	GAA E	AAA K	AAA K	GAG	TTA L
AGG R	GCT A	AAG K	CAA CTG Q L	AAG K	GGC G
AAC N	AAT N	ATC I	CAA Q	GAT AAG D K	${ m TTA}$
GTG V	TTT F	САА	GAG	GAC D	GCC
${ m TTG}$	GAG E	AGA R	AAA K	GCT A	GCT
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Figure 15D

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							gac Asp									240
							gct Ala									288
							caa Gln									336
ccc Pro	cga Arg	ttg Leu	gtg Val	aga Arg	cca Pro	gag Glu	gtt Val	gat Asp	gtg Val	atg Met	tgc Cys	act Thr	gct Ala	ttt Phe	cat His	384

		115					120					125				
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			_		aca Thr		_	_		-	_	-		-	_	528
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					tgc Cys											768
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	_	_	_	_	ttc Phe			_		_		-		_	-	1008
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				cca Pro			_				_					1296
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				cta Leu												1392
~ ~		~		gta Val	_	_	_	_	•		_	_				1440
				cga Arg 485												1488
	_			gag Glu			-									1536
	_			tct Ser		_					_				_	1584
		_		gtg Val			_		_						_	1632
				gat Asp												1680
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Gln Cys Pro Phe Glu Asp His Val Lys Leu Val Asn Glu Val Thr Glu 35 40 45

Phe Ala Lys Thr Cys Val Ala Asp Glu Ser Ala Glu Asn Cys Asp Lys 50 55 60

Ser Leu His Thr Leu Phe Gly Asp Lys Leu Cys Thr Val Ala Thr Leu 65 70 75 80

Arg Glu Thr Tyr Gly Glu Met Ala Asp Cys Cys Ala Lys Gln Glu Pro 85 90 95

Glu Arg Asn Glu Cys Phe Leu Gln His Lys Asp Asn Pro Asn Leu 100 105 110

Pro Arg Leu Val Arg Pro Glu Val Asp Val Met Cys Thr Ala Phe His 115 120 125

Asp Asn Glu Glu Thr Phe Leu Lys Lys Tyr Leu Tyr Glu Ile Ala Arg 130 135 140

Arg His Pro Tyr Phe Tyr Ala Pro Glu Leu Leu Phe Phe Ala Lys Arg 145 150 155 160

Tyr Lys Ala Ala Phe Thr Glu Cys Cys Gln Ala Ala Asp Lys Ala Ala 165 170 175

Cys Leu Leu Pro Lys Leu Asp Glu Leu Arg Asp Glu Gly Lys Ala Ser 180 185 190

Ser Ala Lys Gln Arg Leu Lys Cys Ala Ser Leu Gln Lys Phe Gly Glu 195 200 205

Arg Ala Phe Lys Ala Trp Ala Val Ala Arg Leu Ser Gln Arg Phe Pro 210 215 220

Lys Ala Glu Phe Ala Glu Val Ser Lys Leu Val Thr Asp Leu Thr Lys 225 230 235 240

Val His Thr Glu Cys Cys His Gly Asp Leu Leu Glu Cys Ala Asp Asp 245 250 255

Arg Ala Asp Leu Ala Lys Tyr Ile Cys Glu Asn Gln Asp Ser Ile Ser 260 265 270

Ser Lys Leu Lys Glu Cys Cys Glu Lys Pro Leu Leu Glu Lys Ser His 275 280 285

Cys Ile Ala Glu Val Glu Asn Asp Glu Met Pro Ala Asp Leu Pro Ser

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295

290

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Gln Val Ser Thr Pro Thr Leu Val Glu Val Ser Arg Asn Leu Gly Lys 420 425 430

Val Gly Ser Lys Cys Cys Lys His Pro Glu Ala Lys Arg Met Pro Cys 435 440 445

Ala Glu Asp Tyr Leu Ser Val Val Leu Asn Gln Leu Cys Val Leu His 450 455 460

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485 490 495

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Leu Val Glu Leu Val Lys His Lys Pro Lys Ala Thr Lys Glu Gln Leu 530 540

Lys Ala Val Met Asp Asp Phe Ala Ala Phe Val Glu Lys Cys Cys Lys 545 550 550 555

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Thr Arg Val Glu Pro Pro Gln Tyr Met Ile Asp Leu Tyr Asn Arg Tyr 85 90 95

Thr Ser Asp Lys Ser Thr Thr Pro Ala Ser Asn Ile Val Arg Ser Phe 100 105 110

Ser Met Glu Asp Ala Ile Ser Ile Thr Ala Thr Glu Asp Phe Pro Phe 115 120 125

Gln Lys His Ile Leu Leu Phe Asn Ile Ser Ile Pro Arg His Glu Gln 130 135 140

Ile Thr Arg Ala Glu Leu Arg Leu Tyr Val Ser Cys Gln Asn His Val
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Asp Pro Ser His Asp Leu Lys Gly Ser Val Val Ile Tyr Asp Val Leu 165 170 175

Asp Gly Thr Asp Ala Trp Asp Ser Ala Thr Glu Thr Lys Thr Phe Leu 180 185 190

Val Ser Gln Asp Ile Gln Asp Glu Gly Trp Glu Thr Leu Glu Val Ser 195 200 205

Ser Ala Val Lys Arg Trp Val Arg Ser Asp Ser Thr Lys Ser Lys Asn

210 215 220

Lys Leu Glu Val Thr Val Glu Ser His Arg Lys Gly Cys Asp Thr Leu 225 230 235 240

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Phe Ser Asn Asp His Ser Ser Gly Thr Lys Glu Thr Arg Leu Glu Leu 260 265 270

Arg Glu Met Ile Ser His Glu Gln Glu Ser Val Leu Lys Lys Leu Ser 275 280 285

Lys Asp Gly Ser Thr Glu Ala Gly Glu Ser Ser His Glu Glu Asp Thr 290 295 300

Asp Gly His Val Ala Ala Gly Ser Thr Leu Ala Arg Arg Lys Arg Ser 305 310 315 320

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Ala Tyr Glu Cys Lys Gly Gly Cys Phe Phe Pro Leu Ala Asp Asp Val\$355\$

Thr Pro Thr Lys His Ala Ile Val Gln Thr Leu Val His Leu Lys Phe 370 375 380

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Val Ile Thr Asp Glu Asn Trp Arg Glu Leu Leu Glu Gly Asp Trp Met 35 40 45

Ile Glu Phe Tyr Ala Pro Trp Cys Pro Ala Cys Gln Asn Leu Gln Pro 50 55 60

Glu Trp Glu Ser Phe Ala Glu Trp Gly Glu Asp Leu Glu Val Asn Ile

70 75 65 80 Ala Lys Val Asp Val Thr Glu Gln Pro Gly Leu Ser Gly Arg Phe Ile 90 Ile Thr Ala Leu Pro Thr Ile Tyr His Cys Lys Asp Gly Glu Phe Arg Arg Tyr Gln Gly Pro Arg Thr Lys Lys Asp Phe Ile Asn Phe Ile Ser Asp Lys Glu Trp Lys Ser Ile Glu Pro Val Ser Ser Trp Phe Gly Pro 135 Gly Ser Val Leu Met Ser Ser Met Ser Ala Leu Phe Gln Leu Ser Met 155 150 Trp Ile Arg Thr Cys His Asn Tyr Phe Ile Glu Asp Leu Gly Leu Pro 170 Val Trp Gly Ser Tyr Thr Val Phe Ala Leu Ala Thr Leu Phe Ser Gly 185 Leu Leu Gly Leu Cys Met Ile Phe Val Ala Asp Cys Leu Cys Pro 200 205 Ser Lys Arg Arg Pro Gln Pro Tyr Pro Tyr Pro Ser Lys Lys Leu . 215 Leu Ser Glu Ser Ala Gln Pro Leu Lys Lys Val Glu Glu Glu Gln Glu 230 235 Ala Asp Glu Glu Asp Val Ser Glu Glu Glu Ala Glu Ser Lys Glu Gly 250 Thr Asn Lys Asp Phe Pro Gln Asn Ala Ile Arg Gln Arg Ser Leu Gly 265 Pro Ser Leu Ala Thr Asp Lys Ser <210> 76 <211> 112 <212> PRT <213> Homo sapiens <400> 76 Met Phe Trp Val Met Glu Thr Ala Lys Pro Pro Val Ser Glu Asp Ser Phe Arg Leu Pro Arg Lys Trp Gly Trp Arg Thr Glu Ala Thr Ala Pro His Ala Pro Val Pro Gln Ser Ile Cys Pro Arg Tyr Thr Ser Pro Cys Ala Pro His Asp Cys Gly Ser Gln Thr Val Gln Gly Asn Ser Leu Ser 50

33

Leu Phe Tyr Thr Leu Ser His Lys Ala Pro Gln Leu Pro His Arg Val

65 70 75 80

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n Lys Ala Asp Asp Gly Cys Ser Pro As
n Lys 35 40 45

Met Lys Thr Val Lys Cys Ala Pro Gly Val Asp Val Cys Thr Glu Ala 50 55 60

Val Gly Ala Val Glu Thr Ile His Gly Gln Phe Ser Leu Ala Val Arg
65 70 75 80

Gly Cys Gly Ser Gly Leu Pro Gly Lys Asn Asp Arg Gly Leu Asp Leu 85 90 95

His Gly Leu Leu Ala Phe Ile Gln Leu Gln Gln Cys Ala Gln Asp Arg 100 105 110

Cys Asn Ala Lys Leu Asn Leu Thr Ser Arg Ala Leu Asp Pro Ala Gly 115 120

Asn Glu Ser Ala Tyr Pro Pro Asn Gly Val Glu Cys Tyr Ser Cys Val 130 135 140

Gly Leu Ser Arg Glu Ala Cys Gln Gly Thr Ser Pro Pro Val Val Ser 145 150 155 160

Cys Tyr Asn Ala Ser Asp His Val Tyr Lys Gly Cys Phe Asp Gly Asn 165 170 175

Val Thr Leu Thr Ala Ala Asn Val Thr Val Ser Leu Pro Val Arg Gly
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Cys Val Gln Asp Glu Phe Cys Thr Arg Asp Gly Val Thr Gly Pro Gly
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Phe Thr Leu Ser Gly Ser Cys Cys Gln Gly Ser Arg Cys Asn Ser Asp 210 215 220

Leu Arg Asn Lys Thr Tyr Phe Ser Pro Arg Ile Pro Pro Leu Val Arg 225 230 235 240

Leu Pro Pro Pro Glu Pro Thr Thr Val Ala Ser Thr Thr Ser Val Thr

245 250 255

Thr Ser Thr Ser Ala Pro Val Arg Pro Thr Ser Thr Thr Lys Pro Met 260 265 270

Pro Ala Pro Thr Ser Gln Thr Pro Arg Gln Gly Val Glu His Glu Ala 275 280 285

Ser Arg Asp Glu Glu Pro Arg Leu Thr Gly Gly Ala Ala Gly His Gln 290 295 300

Asp Arg Ser Asn Ser Gly Gln Tyr Pro Ala Lys Gly Gly Pro Gln Gln 305 310 315 320

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<213> Homo sapiens

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Ile Leu Arg Asp Trp Leu Tyr Glu His Arg Tyr Asn Ala Tyr Pro Ser 50 55 60

Glu Gln Glu Lys Ala Leu Leu Ser Gln Gln Thr His Leu Ser Thr Leu 65 70 75 80

Gln Val Cys Asn Trp Phe Ile Asn Ala Arg Arg Arg Leu Leu Pro Asp 85 90 95

Met Leu Arg Lys Asp Gly Lys Asp Pro Asn Gln Phe Thr Ile Ser Arg 100 105 110

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115 120 125

Ile Lys Asn Phe Met Pro Ala Leu Glu Glu Thr Pro Phe His Ser Cys 130 135 140

Thr Ala Gly Pro Asn Pro Thr Leu Gly Arg Pro Leu Ser Pro Lys Pro 145 150 155 160

Ser Ser Pro Gly Ser Val Leu Ala Arg Pro Ser Val Ile Cys His Thr 165 170 175

Thr Val Thr Ala Leu Lys Asp Val Pro Phe Ser Leu Cys Gln Ser Val

180 185 190

Gly Val Gly Gln Asn Thr Asp Ile Gln Gln Ile Ala Ala Lys Asn Phe 195 200 205

Thr Asp Thr Ser Leu Met Tyr Pro Glu Asp Thr Cys Lys Ser Gly Pro 210 220

Ser Thr Asn Thr Gln Ser Gly Leu Phe Asn Thr Pro Pro Pro Thr Pro 225 230 235 240

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Gly Gly Lys Arg Phe Ser His Ser Gly Asn Gln Leu Asp Gly Pro 35 . 40 45

Ile Thr Ala Leu Arg Val Arg Val Asn Thr Tyr Tyr Ile Val Gly Leu 50 60

Gln Val Arg Tyr Gly Lys Val Trp Ser Asp Tyr Val Gly Gly Arg Asn 65 70 75 80

Val Ser Gly Lys Tyr Lys Trp Tyr Leu Lys Lys Leu Val Phe Val Thr 100 105 110

Asp Lys Gly Arg Tyr Leu Ser Phe Gly Lys Asp Ser Gly Thr Ser Phe 115 120 125

Asn Ala Val Pro Leu His Pro Asn Thr Val Leu Arg Phe Ile Ser Gly 130 135

Arg Ser Gly Ser Leu Ile Asp Ala Ile Gly Leu His Trp Asp Val Tyr 145 150 155 160

Pro Thr Ser Cys Ser Arg Cys 165

OR OTHER BIOLOGICAL MATERIAL					
	(90)	CT Rule 13bis)			
A. The indications made below relate to the didescription on page 96, line 30.	eposited nuc	roorganism or o	ther biological material referred to in the	÷	
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet					
Name of depositary institution: Ameri	can Type (Culture Colle	ction		
Address of depositary institution (inclu 10801 University Boulevard Manassas, Virginia 20110-2209 United States of America	ding posta	rl code and co	ountry)		
Date of deposit		Accession Nun	nber		
11 April 2001			PTA-3276		
C. ADDITIONAL INDICATIONS (leave blo	nık ıf not appl	licable)	This information is continued on an addition	nal sheer	
D. DESIGNATED STATES FOR WHICH	INDICATIO	Ons are mai	DE (if the indications are not for all designated St	ales)	
Europe In respect of those designations in which a Europeration of the mention of the grant of withdrawn or is deemed to be withdrawn, only by sample (Rule 28(4) EPC).	of the Europe:	an patent of until	the date on which the application has been	n refused or ting the	
E. SEPARATE FURNISHING OF INDICA	TIONS (leave	e blank if nor applicabl	s)		
The indications listed below will be submitted to the Number of Deposit")	e i nternati onal	i Bureau later (spe	cify the general nature of the indications e.s	"Accession	
For receiving Office use only			For International Bureau use only		
This sheet was received with the international up	phicanon	This sheet	vas received by the International Bureau on	•	
Authorized officer	Authorized officer				
Revised Form PCT/RC//134 (January 2001)		<u> </u>		Penal34co sol6s	

1

PCT/US01/11991

ATCC Deposit No.: PTA-3276

CANADA

The applicant requests that, until either a Canadian patent has been issued on the basis of an application or the application has been refused, or is abandoned and no longer subject to reinstatement, or is withdrawn, the Commissioner of Patents only authorizes the furnishing of a sample of the deposited biological material referred to in the application to an independent expert nominated by the Commissioner, the applicant must, by a written statement, inform the International Bureau accordingly before completion of technical preparations for publication of the international application.

NORWAY

The applicant hereby requests that the application has been laid open to public inspection (by the Norwegian Patent Office), or has been finally decided upon by the Norwegian Patent Office without having been laid open inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Norwegian Patent Office not later than at the time when the application is made available to the public under Sections 22 and 33(3) of the Norwegian Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on the list of recognized experts drawn up by the Norwegian Patent Office or any person approved by the applicant in the individual case.

AUSTRALIA

The applicant hereby gives notice that the furnishing of a sample of a microorganism shall only be effected prior to the grant of a patent, or prior to the lapsing, refusal or withdrawal of the application, to a person who is a skilled addressee without an interest in the invention (Regulation 3.25(3) of the Australian Patents Regulations).

FINLAND

The applicant hereby requests that, until the application has been laid open to public inspection (by the National Board of Patents and Regulations), or has been finally decided upon by the National Board of Patents and Registration without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art.

UNITED KINGDOM

The applicant hereby requests that the furnishing of a sample of a microorganism shall only be made available to an expert. The request to this effect must be filed by the applicant with the International Bureau before the completion of the technical preparations for the international publication of the application.

ATCC Deposit No.: PTA-3276

DENMARK

The applicant hereby requests that, until the application has been laid open to public inspection (by the Danish Patent Office), or has been finally decided upon by the Danish Patent office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the Danish Patent Office not later that at the time when the application is made available to the public under Sections 22 and 33(3) of the Danish Patents Act. If such a request has been filed by the applicant, any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Danish Patent Office or any person by the applicant in the individual case.

SWEDEN

The applicant hereby requests that, until the application has been laid open to public inspection (by the Swedish Patent Office), or has been finally decided upon by the Swedish Patent Office without having been laid open to public inspection, the furnishing of a sample shall only be effected to an expert in the art. The request to this effect shall be filed by the applicant with the International Bureau before the expiration of 16 months from the priority date (preferably on the Form PCT/RO/134 reproduced in annex Z of Volume I of the PCT Applicant's Guide). If such a request has been filed by the applicant any request made by a third party for the furnishing of a sample shall indicate the expert to be used. That expert may be any person entered on a list of recognized experts drawn up by the Swedish Patent Office or any person approved by a applicant in the individual case.

NETHERLANDS

The applicant hereby requests that until the date of a grant of a Netherlands patent or until the date on which the application is refused or withdrawn or lapsed, the microorganism shall be made available as provided in the 31F(1) of the Patent Rules only by the issue of a sample to an expert. The request to this effect must be furnished by the applicant with the Netherlands Industrial Property Office before the date on which the application is made available to the public under Section 22C or Section 25 of the Patents Act of the Kingdom of the Netherlands, whichever of the two dates occurs earlier.

INDICATIONS BELATING TO A DEPOSITED MICROORGANISM

OR OTHER BIOLOGICAL MATERIAL						
(PCT Rule 13bis)						
A. The indications made below relate to the deposited microorganism or other biological material referred to in the description on page 96, line 30.						
B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet 🗵						
Name of depositary institution: American T	ype Culture Collection					
Address of depositary institution (including p 10801 University Boulevard Manassas, Virginia 20110-2209 United States of America	postal code and country)					
Date of deposit	Accession Number					
11 April 2001	PTA-3277					
C. ADDITIONAL INDICATIONS (leave blank if no	rapplicable) This information is continued on an additional sheer					
D. DESIGNATED STATES FOR WHICH INDIC	ATIONS ARE MADE (of the indications are not for all designated States)					
until the publication of the mention of the grant of the E	tent is sought a sample of the deposited microorganism will be made available uropean patent or until the date on which the application has been refused or such a sample to an expert nominated by the person requesting the Continued on additional sheets					
E. SEPARATE FURNISHING OF INDICATION	S (leave blank if not applicable)					
The indications listed below will be submitted to the international Number of Deposit")	ational Bureau later (specify the general nature of the indications e.g. "Accession					
For receiving Office use unly	For International Bureau use only					
☐ This sheet was received with the international application	on This sheet was received by the International Bureau on					
Authorized officer	Authorized officer					
Revised Form PCT/RO/134 (January 2001)	Parrol Saen volte					

INTERNATIONAL SEARCH REPORT

International application No. PCT/US01/11941

	SIFICATION OF SUBJECT MATTER						
` '	A61K 37/02; C12N 15/00						
	US CL: 550/350; 485/7.1 According to International Patent Classification (IPC) or to both national classification and IPC						
B. FIEL	DS SEARCHED						
Minimum do	ocumentation searched (classification system followed	by classification symbols)					
	530/350; 435/7.1						
Documentati	on searched other than minimum documentation to	the extent that such documents are in	ncluded in the fields				
seaEchred GE	NEMBL, AGENESEQ						
Electronic d	ata base consulted during the international search (na	ame of data base and, where practicable	e, search terms used)				
STN: ME	DLINE, BIOSIS, USPAT, JAPIO, HCAPLUS						
C. DOC	UMENTS CONSIDERED TO BE RELEVANT						
 	Citation of document, with indication, where app	was into a fith a relevant pagagons	Relevant to claim No.				
Category*	Citation of document, with indication, where app	ropriate, of the relevant passages					
X	WO 95/23857 A1 (DELTA BIOTEC	HNOLOGY LIMITED) 08	1-9, 15-19				
	September 1995 (08.09.95), see entire	document.					
	_						
X	EP 0 322 094 A1 (DELTA BIOTECHN	OLOGY LIMITED) 28 June	1-9, 15-19				
1	1989 (28.06.89), see entire document.						
		:	,				
		·					
Furth	ner documents are listed in the continuation of Box C						
1	CIMI ORNOROLLOS OL CINCA DOCUMENTA	"I" later document published after the into date and not in conflict with the app	lication but cited to understand				
	nment defining the general state of the art which is not considered be of particular relevance	the principle or theory underlying the					
"E" ean	lier document published on or after the international filing date	"X" document of particular relevance; the considered novel or cannot be considered.					
"L" doc	ument which may throw doubts on priority claim(s) or which is do to establish the publication date of another citation or other	when the document is taken alone					
spe	cial reason (as specified)	"Y" document of particular relevance; the considered to involve an inventive step	when the document is combined				
	nment referring to an oral disclosure, use, exhibition or other	with one or more other such docum obvious to a person skilled in the art	nents, such combination being				
"P" doc	nument published prior to the international filing date but later on the priority date claimed	"&" document member of the same patent	family				
		Date of mailing of the international se	arch report				
		Date of mailing of the international se	G 7001				
18 JULY	2001						
Name and r	nailing address of the ISA/US	Authorized officer ve	AMMA				
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Authorized officery HOBE ROBINSON							
1	1, D.C. 20231	Telephone No. (703) 308-0196	i 15°				
Facsimile N	o. (703) 305-3230	1 00 000-0100	1/				

INTERNATIONAL SEARCH REPORT

International application No. PCT/US01/11991

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)					
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:					
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:					
2. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:					
3. X Claims Nos.: 10-14, 20-32 and 34-36 because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).					
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)					
This International Searching Authority found multiple inventions in this international application, as follows:					
Please See Extra Sheet.					
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.					
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.					
S. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:					
4. X No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-9, 15-19 (FGF-7)					
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.					

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

Group 39, all partially as in Group 1, concerning Therapeutic Protein X: TR12.

Group 40, all partially as in Group 1, concerning Therapeutic Protein X: TR13.

Group 41, all partially as in Group 1, concerning Therapeutic Protein X: TR14.

Group 42, all partially as in Group 1, concerning Therapeutic Protein X: TR16.

Group 43, all partially as in Group 1, concerning Therapeutic Protein X: HLDOU18.

Group 44, all partially as in Group 1, concerning Therapeutic Protein X: HSDSB09.

Group 45, all partially as in Group 1, concerning Therapeutic Protein X: HDPBQ71.

Group 46, all partially as in Group 1, concerning Therapeutic Protein X: HAGDG59.

Group 47, all partially as in Group 1, concerning Therapeutic Protein X: HCHNF25.

Group 48, all partially as in Group 1, concerning Therapeutic Protein X: HKACD58.

Group 50, all partially as in Group 1, concerning Therapeutic Protein X: HWACB86.

Group 51, all partially as in Group 1, concerning Therapeutic Protein X: HRDFD27.

Group 52, all partially as in Group 1, concerning Therapeutic Protein X: HRDFD27.

Group 53, all partially as in Group 1, concerning Therapeutic Protein X: HCEGG08.

Group 53, all partially as in Group 1, concerning Therapeutic Protein X: HKACI79.

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This ISA found multiple inventions as follows:
Groups 1-58, claim(s) 1-9, 15-19, all in part, drawn to a therapeutic protein X, wherein X correlates to those listed in
the Table on page 11 of the description. If any of Groups 1-58 are elected the claims will only be examined in-so-far-
as it pertains to the elected protein X. For example,
If Group 1 is elected, this correlates to Therapeutic Protein X:FGF-7.
Group 2, all partially as in Group 1, concerning Therapeutic Protein X: Kertainocyte growth factor 2.
Group 3, all partially as in Group 1, concerning Therapeutic Protein X: C-C Chemokine Receptor 5.
Group 4, all partially as in Group 1, concerning Therapeutic Protein X: Cathepsin K.
Group 5, all partially as in Group 1, concerning Therapeutic Protein X: MPIF-1 (Myeloid Progenitor Inhibitory
Factor).
Group 6, all partially as in Group 1, concerning Therapeutic Protein X: VEGF.
Group 7, all partially as in Group 1, concerning Therapeutic Protein X: VEGF-2.
Group 8, all partially as in Group 1, concerning Therapeutic Protein X: BLyS.
Group 9, all partially as in Group 1, concerning Therapeutic Protein X: KD1.
Group 10, all partially as in Group 1, concerning Therapeutic Protein X: TMP-1.
Group 11, all partially as in Group 1, concerning Therapeutic Protein X: TMP-2.
Group 12, all partially as in Group 1, concerning Therapeutic Protein X: TMP-3.
Group 13, all partially as in Group 1, concerning Therapeutic Protein X: TMP-4.
Group 14, all partially as in Group 1, concerning Therapeutic Protein X: Connective Tissue Growth Factor Protein.
Group 15, all partially as in Group 1, concerning Therapeutic Protein X:CTGF-2.
Group 16, all partially as in Group 1, concerning Therapeutic Protein X: Connective Tissue Growth Factor 4.
Group 17, all partially as in Group 1, concerning Therapeutic Protein X: Human T-cell Lymphoma-lipoprotein
associated phospholipase-A2.
Group 18, all partially as in Group 1, concerning Therapeutic Protein X: VEGI.
Group 19, all partially as in Group 1, concerning Therapeutic Protein X: AIM-1.
Group 20, all partially as in Group 1, concerning Therapeutic Protein X: TNF-delta.
Group 21, all partially as in Group 1, concerning Therapeutic Protein X: TNF-epsilon.
Group 22, all partially as in Group 1, concerning Therapeutic Protein X: AIM-2.
Group 23, all partially as in Group 1, concerning Therapeutic Protein X: Endokine.
Group 24, all partially as in Group 1, concerning Therapeutic Protein X: TR1.
Group 25, all partially as in Group 1, concerning Therapeutic Protein X:TR2.
Group 26, all partially as in Group 1, concerning Therapeutic Protein X: DRs.
Group 27, all partially as in Group 1, concerning Therapeutic Protein X: TR4.
Group 28, all partially as in Group 1, concerning Therapeutic Protein X: 4-1BBsv receptor.
Group 29, all partially as in Group 1, concerning Therapeutic Protein X: OPGL.
Group 30, all partially as in Group 1, concerning Therapeutic Protein X: FasL.
Group 31, all partially as in Group 1, concerning Therapeutic Protein X: Fas.
Group 32, all partially as in Group 1, concerning Therapeutic Protein X: TR5.
Group 38, all partially as in Group 1, concerning Therapeutic Protein X: TR6.
Group 34, all partially as in Group 1, concerning Therapeutic Protein X: DR5.
Group 35, all partially as in Group 1, concerning Therapeutic Protein X: TR8.
Group 36, all partially as in Group 1, concerning Therapeutic Protein X: TR9.
Group 37, all partially as in Group 1, concerning Therapeutic Protein X: TR10.
Group 38, all partially as in Group 1, concerning Therapeutic Protein X: TR11.
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INTERNATIONAL SEARCH REPORT

International application No. PCT/US01/11991

Group 54, all Group 55, all Group 56, all	partially	as in	Group	ı,	concerning	Therapeutic	Protein	\mathbf{X} :	HDTAI21.	
Group 57, all Group 58, all										

Group 59-117, claim(s) 33, drawn to a method of extending shelf life of Therapeutic Protein X, wherein Therapeutic Protein X can be any of the proteins listed in the Table on page 11 of the description. If any of Groups 59-117 are elected the claim will only be examined in-so-far-as it pertains to the elected Protein X.

The inventions listed as Groups 1-118 do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: because the technical feature linking groups 1-118 is not special because Inventions 1-58 do not avoid the prior art as Delta Biotechnology Limited, (EP 322094, June 28, 1989) teaches the claimed sequence. Thus, the invention does not relate to a single inventive concept and is not a contribution over the prior art.